

## Biosynthesis of mesoporous silica supported silver nanoparticles for antibacterial evaluation

SIDRA GHAFOOR<sup>1a\*</sup>

<sup>1</sup>Research Scholar department of Chemistry, University of Sialkot Punjab, Pakistan

<sup>2</sup>Postdoctoral Fellow, Department of Applied Physics, The Hong Kong Polytechnic University, Hong Kong.

Corresponding Email: <sup>a</sup>[sidraghafoorofficial@gmail.com](mailto:sidraghafoorofficial@gmail.com), <https://orcid.org/0000-3450-1824-4956-2>

### Abstract

Nanotechnology's game-changing applications have boosted the importance of synthesizing nanomaterials with carefully controlled dimensions. Biosynthesis of nanomaterials is a developing field that seeks to synthesize various nanoparticles. Leaf extract from *Azadirachta indica* will be used to bio generate silver nanoparticles on mesoporous silica spheres for this study. The leaf extract will be processed using ethanol. A modified version of Stober's

method will be used to synthesize mesoporous silica balls. Spectroscopy, x-ray diffraction, scanning electron microscopy, and other cutting-edge tools will be employed to characterize the produced silica supported silver nanoparticles. The disc diffusion method or the Agar well diffusion method will be used to evaluate the antibacterial efficacy of the synthesized nanoparticles.

**KEYWORD:** (TEOS) Tetra ethyl orthosilicate, (TMB) Trimethyl benzene, (CTAB) Hexadecyl trimethylammonium bromide, (MSNs) mesoporous silica nanospheres, (NPs) nanoparticles, (FTIR) Fourier transform infrared spectroscopy, (UV VISI) Ultra violet, (XRD) X-Ray diffraction, (SEM) Scanning electron microscopy

### INTRODUCTION

The fundamental principle of Nano science is that material properties are dimension-dependent. Although "nano science" and technology have been around for a while, they are rapidly developing subjects that are causing revolutions in all sciences on par with what genomics and proteomics did lately for the biological sciences. The size of a nanosac, or approximately  $10^{-9}$  m is the basis for the use of the prefix Nano (Hochella Jr., 2002).

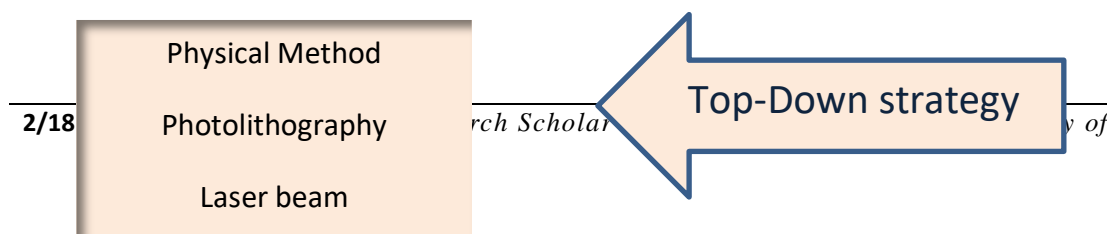
## BIOSYNTHESIS OF MESOPOROUS SILICA SUPPORTED SILVER NANOPARTICLES FOR ANTIBACTERIAL EVALUATION

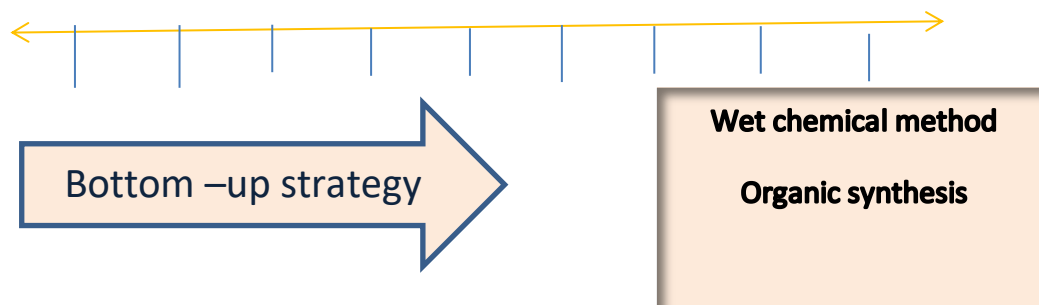
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The diminutive Latin term from which "Nano" is derived. One nanometer is one thousand millionth of a metre (1nm=10 to the ninth power) (Rangasamy, 2011).

One of Nano science's most important uses is in nanotechnology. Definition: Nanotechnology is the study and application of materials with dimensions of 100 nm or less (Mody et al., 2010). The medical sciences find significant use for nanotechnology. Exponential increase in recent years has attested to its significance in the fight against cancer. Nanotechnology-created nanoparticles provide significant untapped potential in the realm of therapeutic cancer development due to their adaptability in terms of design and tuning (Misra et al., 2010).

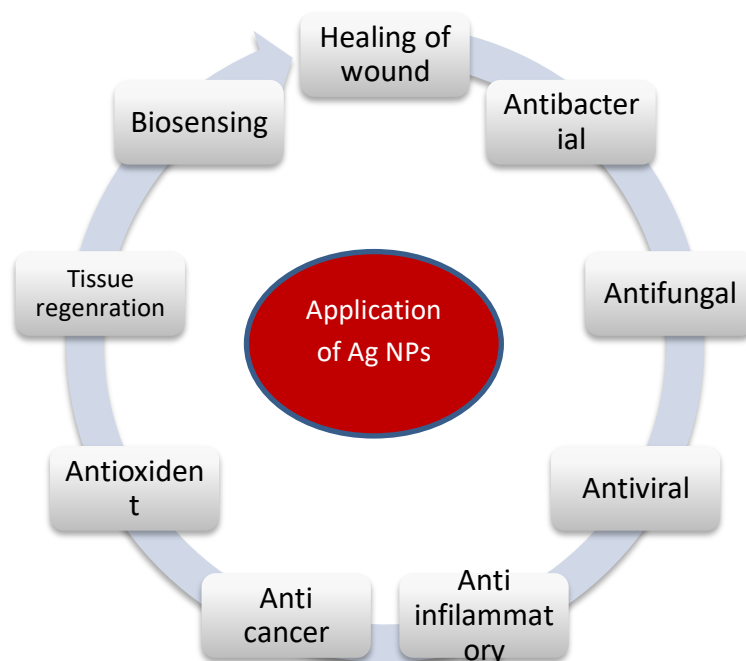
Both methods are used in the synthesis of nanomaterials in nanotechnology. The two most prevalent methods for creating nanoparticles are the top-down and bottom-up approaches. The **"top down"** method involves dismantling macroscale structures into smaller ones. Particles on the order of a few microns in size can be made using this method. If you want to create a certain type of nanomaterial, you can use a top-down method, which is a simpler tool that relies on either separating the bulk material or simplifying the creation process. The lack of surface structure is the main issue with top-down approaches. Nanowires fabricated via the top-down method, for instance, are rough and riddled with surface contaminations and structural faults on the nanoparticles' outer layers. High-energy wet ball milling, electron-beam lithography, gas phase condensation, aerosol, and spray are all examples of such processes. The term **"bottom up approach"** is used to describe the method by which complex Nano structures are built from their component atoms and molecules. Means to construct something from the ground up, either atomically, molecularly, or cluster-by-cluster. Bottom-up methods for the synthesis of luminescent nanoparticles have been widely reported, and include, among others, all organometallic chemical route, reverse micelle route, sol-gel synthesis, and colloidal precipitation, template, assisted sol-gel electrodeposition, etc.





### Use of Bottom Up and Top Down Approach

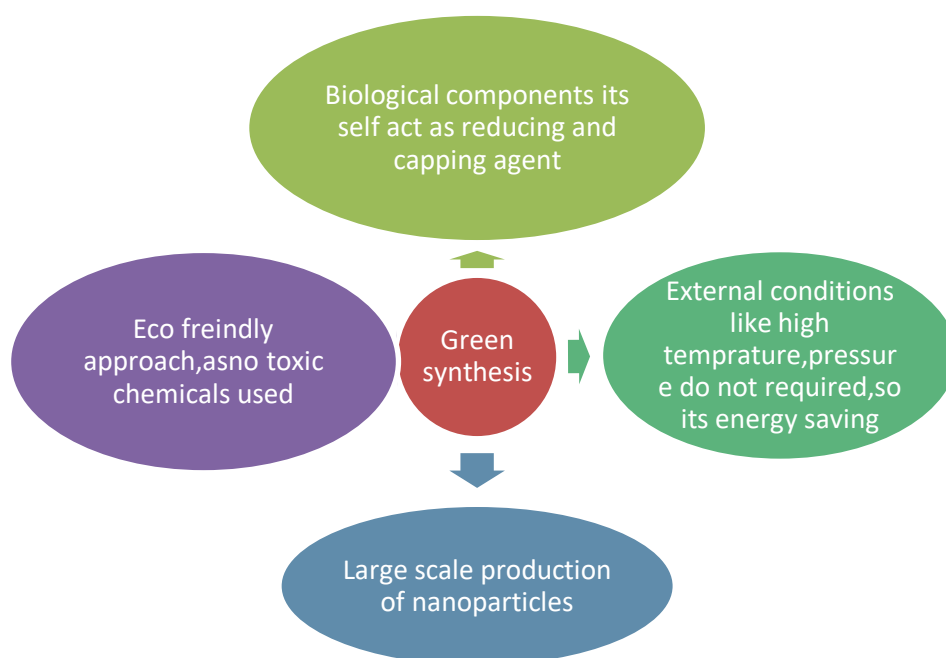
Because nanoparticles are so versatile, we'll go over each possible application in detail below. Because of their magnetic, optical, and chemical characteristics, ferrite nanoparticles are practically ubiquitous. The medical and manufacturing fields can both benefit from using nanoparticles. Their applications range from the biomedical to the technological, including water purification and catalysis. Most commonly, nanoparticles facilitate current flow in electrical chips used in circuits (Kolahalam et al., 2019).



**Fig 1.2: Application of Ag NPs**

A "Green synthesis" refers to the production of nanoparticles from renewable resources like flavonoids and terpenoids. When present in the extract, these terpenoids and flavonoids serve

as a capping and reducing agent. Although conventional techniques have been in use for quite some time, recent research has demonstrated that green techniques are superior for producing NPs due to lower failure risks, lower costs, and easier characterization. Physical and chemical ways of producing NPs have imposed numerous stresses on the environment as a result of their toxic consequences. The synthesis of NPs from plants is undeniably straightforward; in only a few minutes to a few hours, a metal salt can be prepared at room temperature using plant extract. Modern society has come to value plant extract for its vitamin and amino acid content (Gour & Jain, 2019).



**Figure 1.3: Green Synthesis of nanoparticles**

The meliaceae (mahogany) family is where neem calls home. *Azadirachta indica* is a plant with a long history of use as medicine in India and neighbouring countries. It has been recognised for over 200 years as one of the most adaptable medicinal plants available. Likewise, there is a broad spectrum of biological details. Each and every part of the tree can be used to create effective home medicines for a wide range of common human ailments and historical ills. Silver and gold nanoparticles have been synthesised using *Azadirachta indica* leaf extract. The antibacterial activity of biosynthesised silver nanoparticles may be increased when capped with the neem leaf extract, which is the plant's most notable property (Lalitha et al., 2013).

To stabilise the nanoparticles, neem contains phytochemicals such terpenoids and flavanones, which act as reducing and capping agents. Neem leaf extract converts silver salt into

nanoparticles (AgNPs). The neem extract coating gives the produced nanoparticles superior antibacterial action. According to (Verma & Mehata, 2016).

The anti-inflammatory, anti-pyretic, anti-fungal, anti-gastric ulcer, hypoglycemic, diuretic, anti-bacterial, anti-malarial, anti-pandemic, etc. properties of Neem plant leaves have led to its widespread use. The Ayurvedic term for "curing ailment" is "SarvaRogha-Nirvarini," and neem (Nimba) has been used for centuries to treat a. The US National Academy of Sciences has dubbed this worldwide system of problem-solving "The Tree of Problem Solving" (Senthilkumar et al., 2018).

### **1.9 Metallic nanoparticles**

Nanoparticles of metal, often called microparticles, are a type of substance with special qualities that makes it useful in many fields, including business, agriculture, and the home. New MNPs have been rapidly created and mass-produced to fulfil the expanding demand for nanotechnology-based products and gadgets. Reference: (Peralta-Videa et al., 2016).

Green chemistry on mesoporous silica spheres for the creation of silver nanoparticles is the most dependable technology currently available. For the synthesis of silver nanoparticles mesoporous silica nano spheres are used due to their large surface area and higher porosity, and bio functionality of pores.

The stober technique creates mesoporous silica spheres, which can be used to make both silica and non-silica particles. Stobber process actually called the sole gel process in which the hydrolysis and condensation of salinol group in a different solutions are used. MSNs synthesized by this process use tetraorthoosylane, tetraethyl orthosilicates (TEOS), surfactant and solvent. The antibacterial activity of silver NPs generated through biosynthesis is highest. Synthesized silver NPs lose some of their useful properties due to their tendency to aggregate. Therefore, we prepare the nanoparticles on MSN to prevent the aggregation. Nanoparticles' functional qualities are improved thanks to the mesoporous silica sphere's ability to prevent aggregation (Mehmood et al., 2017).

## **RESEARCH METHODOLOGY**

### **3.1. Materials**

Ethanol (>98%), Tetraethyl orthosilicate (>98%), ammonia solution in water (28-30%), (CTAB), trimethyl benzene (TMB), silver nitrate, neem plant extract, NaOH solution to

maintain basic pH, and HCl solution to maintain acidic pH will be used to synthesise silica-supported silver nanoparticles.

### **3.2. Plant collection**

The university of Sialkot's Botany department confirmed in March 2023 that samples of *Azadirachta indica* leaves collected from several locations in Narrowal District, Narrowal, Pakistan were indeed authentic.

### **3.3. Extraction of plant**

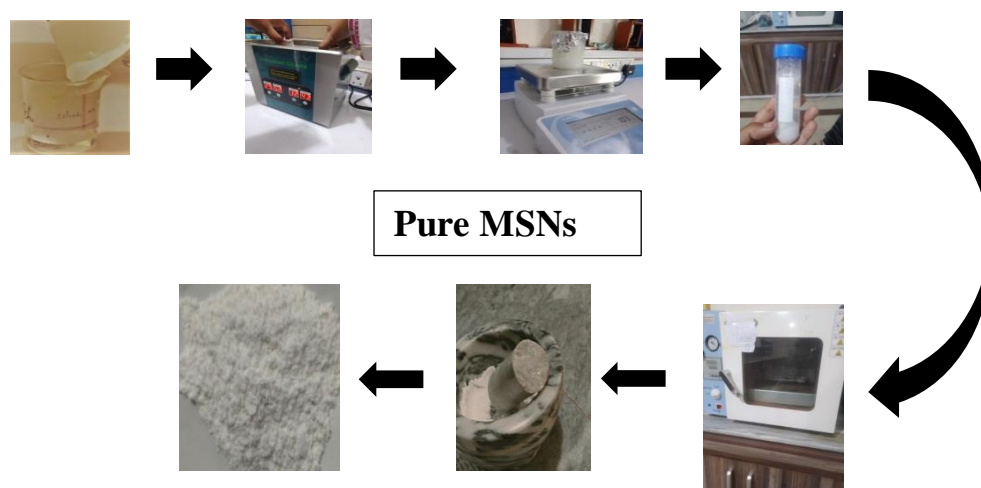
The surface impurities were washed away from the collected *Azdiracta indica* leaves by first washing them in regular tap water and then in deionized water. After being cleaned and dried, the leaves were ground into a powder. The quantity of fine powder was measured, and it came in at 95 grammes. One half of the fine powder weighed 50 grams, while the other half weighed 45 grams. To a container holding 300 millimeter of ethanol, 50 grammes of leaf extract was added. Then, for 30 minutes, the mixture was sonicated at room temperature to ensure uniform blending. The liquid was sonicated, then transferred to a stirring machine and heated to 60 degrees Celsius for thorough mixing. Wattsman No. 1 filter paper was used to filter the mixture. The rotary evaporator was then used to remove the solvent from the final product. A jelly-like result was formed after the solvent was removed; this was then dried in a hot plate to produce powder at the appropriate temperature. The process resulted in a solid extract form. The extracted material was kept cold (4 degrees Celsius) for further use

### **Preparation of mesoporous silica spheres by stobber process**

"Modifications to the stobber method were used to create mesoporous silica spheres. One hundred millilitres of the beaker were removed, rinsed in distilled water, ethanol, and air dried. Then, 16 ml of water and 4 ml of ethanol were added to the beaker, bringing the total volume to 20 ml. 0.6g of CTAB, a white powder surfactant, were included into the mixture. After covering the mixture with aluminum foil, it was sonicated for 15 minutes at room temperature to ensure even distribution of ingredients. A clear solvent was obtained using sonication. The PH of the clear solvent was kept between 12 and 13 with the addition of 0.5 milliliters of a 30% ammonia solution. After making sure the PH was stable, 0.9 ml of trimethyl benzene and 2 ml of tetraethyl orthosilicates were added all at once. Two hours were spent stirring the solution. After 2 hours of constant stirring, the solution thickened noticeably. For 15 minutes, the mixture was spun at 8000 RPM in a centrifuge. White precipitates settled to the bottom of the centrifuge tubes after the process. Precipitates were collected and cleaned with ethanol after

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the solvent in the tube was removed. After centrifuging the solution for 5 minutes, it was shook vigorously to remove any sediment from the bottom of the tube. This cycle of washing was done three times. The resulting precipitates were washed for a third time, then dried in an oven at 120 °C for 12-16 hours. The MSN hardened into a solid state after being exposed to air, and was further powdered using a mortar and pestle. Once the fine powder was collected, it was placed in the tube for later use.



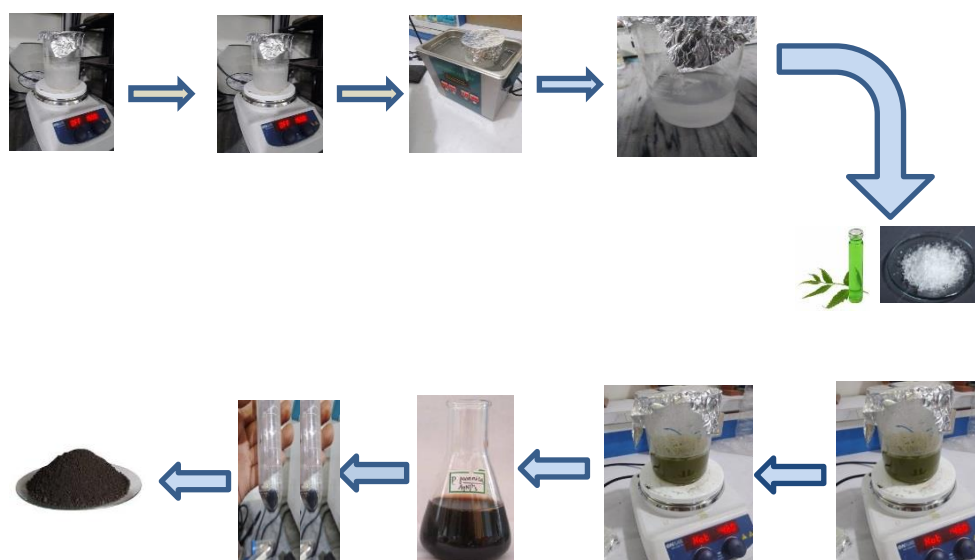
**Synthesis of pure MSNs using stobber process**

### **Biosynthesis of silver Nanoparticles**

A solution of 0.2 gram of mesoporous silica nanospheres (MSN) in 200 milliliters of distilled water was used to synthesize silver nanoparticles. The 0.2g of MSN was added, and after 15 minutes of stirring, the mixture was transferred to a sonicator to be thoroughly mixed at room temperature. The acidity of the solution was kept in the 2-to-3 range to provide uniform dispersion. The required pH was kept at with an HCL solution. The solution became transparent after adding 2 cc of HCL, showing that the MSN was evenly distributed throughout the liquid. The pH was kept constant at 2.8 through constant stirring, and the mixture was thoroughly mixed at all times. A precursor of 1mm silver nitrate (0.034) was added after MSN dispersion. The plant extract was then added, which was diluted in 25 ml of ethanol, and the mixture was kept on the stove and constantly stirred. After that, the mixture was boiled for 5 minutes. Silver nanoparticles formed after the color of the solution changed from dull yellow to dark brown during the boiling process. The initial pH of the solution was low, but a more basic pH range was required for the formation of silver nanoparticles. NaOH solution was made by dissolving

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0.4g of NaOH into 10 ml of distilled water and mixing until the salt was completely dissolved. The pH was adjusted by adding 2 ml of NaOH solution to keep the range of the solution between 8 and 9. The pH was tested and reported as 8 after the addition of NaOH solution. All the while, the solution was being stirred constantly and then allowed to cool to room temperature. The solution was then transferred to the centrifuge tube and spun for 15 minutes. To eliminate any residual chemicals and biomolecules, the resultant product was washed three times with ethanol before being isolated. The product was then dried in an oven at 120 degrees Celsius for a whole day. Once the silver nanoparticles were obtained,



### Biosynthesis of silver nanoparticles using plant extract

## RESULTS AND DATA ANALYSIS

### UV-Vis spectroscopy

Analysing the UV-visible absorption spectrum provides insight into the electrical nature of the optical band gap in the material. Absorption in the near UV spectrum occurs when electronic changes inside the sample are in sync with those outside. Energy in the band gap and UV absorption in SNPs are connected. The UV absorption edge provides a reliable approximation of the band gap for any system.

For the first characterization of the synthesised nanomaterials, UV-visible spectroscopy is a very beneficial instrument. In order to study the synthesis and permanancy. Many assistances are related with its use, such as being time effective, simple, and definite to various kinds of

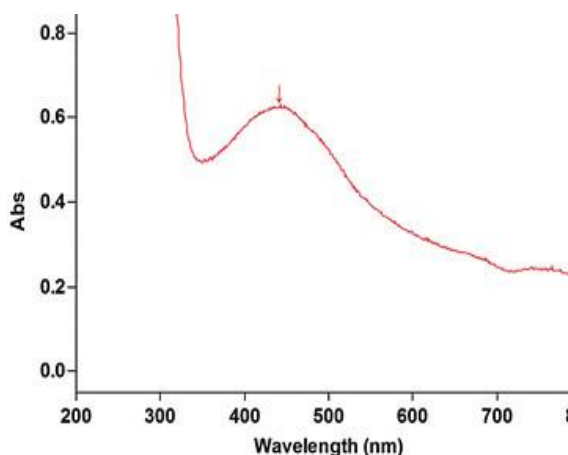


NPs. It is also significant to particle characterization of colloidal suspension of AgNPs, UV-vis has been used. In AgNPs, the conduction and valence bands seem to be quite near to one another, allowing electrons to transfer about easily and produce an SPR absorption band. The size of the particles, the dielectric medium, and the chemical environment all have an effect on how much AgNP absorbs. Surface plasmon peaks are used to capacity the quantity of metal NPs, which range in size from 2 to 100 nm. SPR peaks in the visible region provide as indication of the occurrence of NPs.

According to a research by researcher, the normal peak for AgNPs was seen about 430 nm. Furthermore, dynamic light scattering (DLS) was used to evaluate the nanoparticles' average particle size, which was appeared to be 58.05 nm.

UV visible spectra for silver nanoparticles was plotted using origin software. Graph was plotted between wavelength and absorbance, only one peak obtained at 430 nm. At this place peaks for silver nanoparticles appeared and 430 nm place give the presence of SNPs.

In a work by researcher very unchanging spherical AgNPs were shaped using a green method. A discrete band between 420 and 430 nm was originate in this study's use of UV-visible spectroscopy to look at the stability of nanoparticles over three months. In a succeeding green examination, researcher stated the synthesis of non-spherically shaped AgNPs produced two distinct peaks at 363 and 426 nm.



**Figure 4.1: UV visible spectra of synthesized silver nanoparticles**

#### **XRD analysis of samples.**

X-ray diffraction (XRD) allows for the exact measurement of the size distribution of nanomaterials and the measurement of millions of crystals. X-ray diffraction (XRD) is a common technique for characterising NPs. X-ray diffraction (XRD) is commonly used to

determine the lattice parameters, phase, crystalline grain size, and crystalline structure. The XRD graph plotted using the following settings : The crystal structure of samples was evaluated using X-Ray diffraction technique. The XRD data was

Collected using a portable table top XRD machine.

Data was collected using following settings:

X-Ray source: Copper (Cu) tube with 1.54184(A), Scanning rate : 5 degree per min.

The XRD spectra was plotted using origin software .The crystal structure of synthesized nanoparticles and their crystallite size were calculated using origin software.

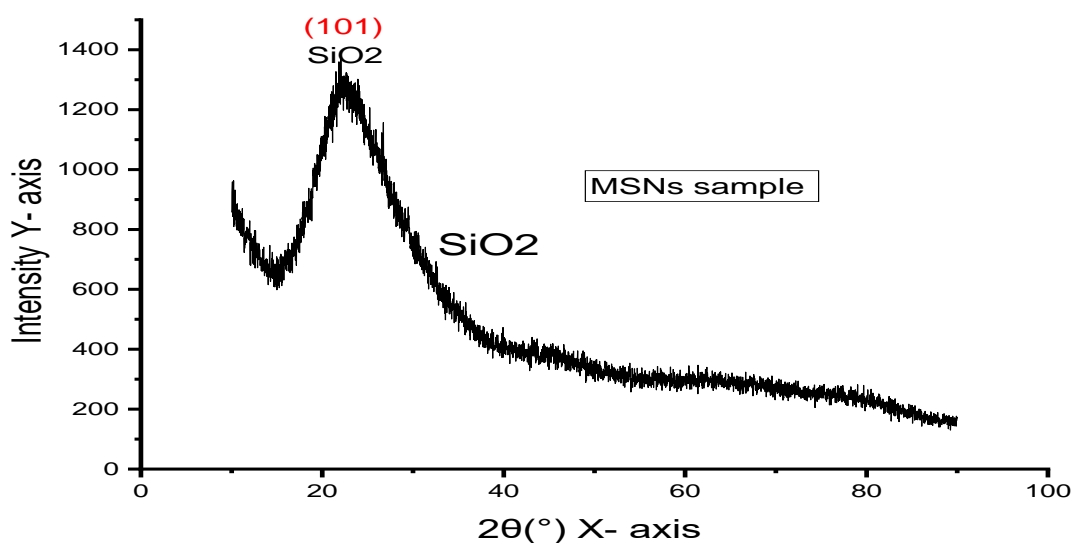
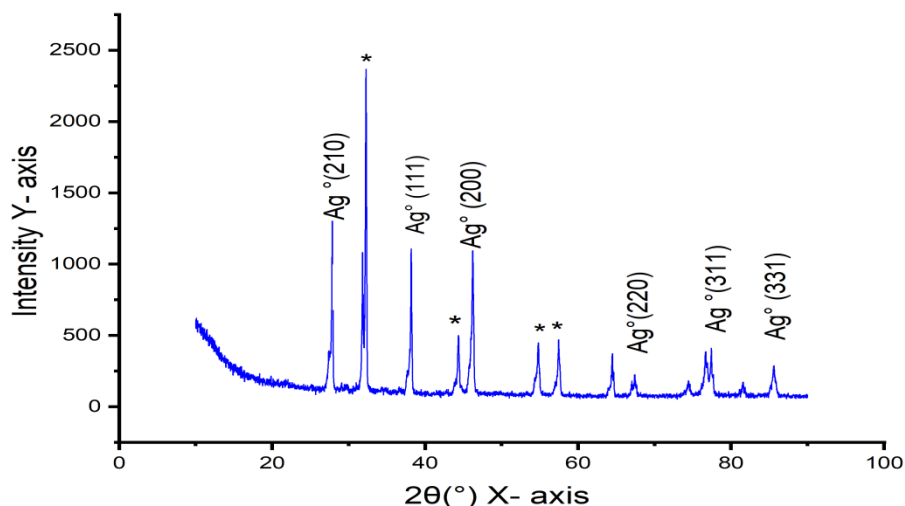


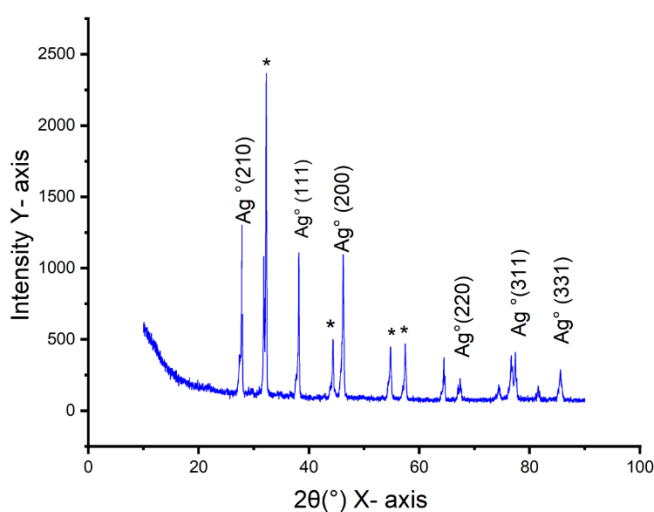
Figure : 4.2: **Determine the pure MSNs presence of silica in the sample.**

The graph was plotted 2 theeta on X – axis against intensity on y- axis a curve is obtained which contain the only one peak at 21 theeta value and crystal lattice value (101) determine the presence of silica SiO2. This determine the presence of silicon dioxide.



**Figure 4.4 : XRD pattern of sample 1 to determine the crystal structure and peaks obtained at (210), (110), (200), (220), (311), (331)**

We have plotted the XRD graph using the origin software. In the graph we have plotted  $2\theta$  value on X-axis and intensity (au) in Y-axis. We obtained the peaks for silver nanoparticles. Six peaks were obtained for pure silver, and the peaks represented by a star indicate impurities in the sample. The impurity belongs to some silver chloride atoms present in the sample. The 1<sup>st</sup> peak was obtained at  $27^\circ$  with hkl value (210), the 2<sup>nd</sup> peak appeared at  $38^\circ$  with hkl value (110), the 3<sup>rd</sup> peak was obtained at (200) at  $46^\circ$ . Other peaks appeared at  $67^\circ$ ,  $77^\circ$ ,  $85^\circ$  with hkl values (220), (311), (331). Other peaks presented with a star belong to impurities present in the sample.

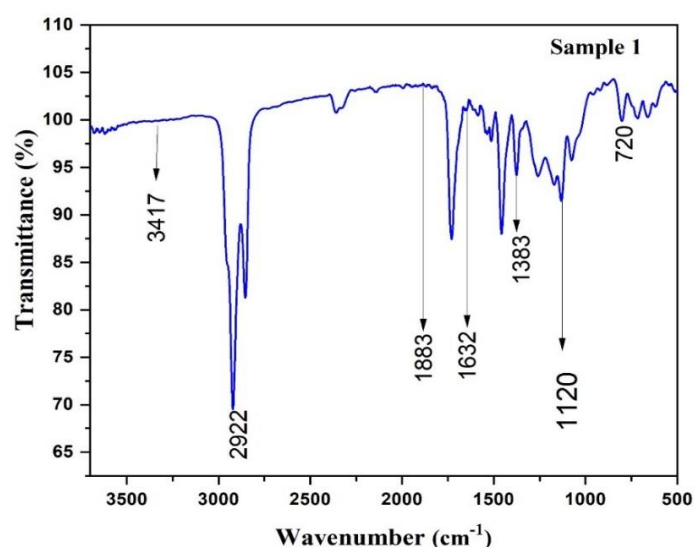


**Figure 4.3: Determine the crystal structure of silver nanoparticles (210).(110),(200).(220),(311),(331).**

We have plotted the XRD graph using the origin software. In the graph we have plotted  $2\theta$  value on X-axis and intensity (au) in Y-axis. We obtained the peaks for silver nanoparticle. 6 peaks obtained for the pure silver and the peaks that are represented by star indicate the impurities in the sample. The impurity belongs to some silver chloride atom present in the sample. 1<sup>st</sup> peak was obtained at  $27^\circ$  with hkl value (210), 2<sup>nd</sup> peak was appeared at  $38^\circ$  with hkl value (110), 3<sup>rd</sup> peak obtained at (200) at  $46^\circ$ . Other peaks were appeared at  $67^\circ$ ,  $77^\circ$ ,  $85^\circ$  with hkl value (220),(311),(331). Other peaks that are presented with star belong to impurities present in the sample.

**FTIR analysis of samples.**

This study's results demonstrated that the synthesized AgNPs controlled functional groups, and that distinguished peaks could be found at 1120, 1383, 1883, 720, 1632, and 3416  $\text{cm}^{-1}$ , corresponding to primary alcohols and  $\text{NO}_3$ , as well as the C–N stretching vibrations of aromatic and aliphatic amines, respectively. The scientists in this study made green nanoparticles from silver nanoparticles and analyzed those using FTIR after preparing green nanoparticles from *Trigonella foenum-graecum*. The findings exposed distinguished bands of absorption for both AgNPs and seed extracts of *Trigonella foenum*. The study's peaks were set at about 3264, 1636, and 1961  $\text{cm}^{-1}$ , which stand for –OH, NH, and =C–H, correspondingly.



Experimental conclusions from FTIR spectroscopy recommended that these results might be matching to those that have been studied and debated away in the context of the green path synthesis of silver nanoparticles, mainly in rights related to microbiology. FTIR spectra of silver nanoparticles was plotted by using origin software. The plotted graph contain the peaks at 3417, 2922, 1632, 1883, 1383, 1120, 720.

### **Antibacterial activity of silver nanoparticles**

In the University of Sialkot's biotechnology lab, the antibacterial activity of produced silver nanoparticles was evaluated using the agar disk diffusion method. *S. aureus* and *Escherichia coli* bacteria's antibacterial activity against each compound. In test tubes, pure cultures of bacteria were introduced to L.B. broth (PH=7) and left to develop for a full day at 37°C. The bacterium suspensions were made with a sterile, isotonic solution. The next step involved creating a 9:1 dilution of bacteria in a test tube by mixing 1 mL of turbid bacterial culture with 9 mL of distilled water.

### **Preparation of Nutrient Agar Medium**

Agar solution was prepared by dissolving 6.25g of nutritional agar powder in 250mL of distilled water. To dissolve them, fully combine them. It was sterilised in an autoclave set at 120 °C for 15 minutes with the pH maintained at 7. After autoclaving, all of the experiments were carried out in a laminar flow. Pour the liquid medium into sanitized Petri plates after allowing it to settle. A clean environment should be used to prepare agar in order to prevent contamination. After it has set, the agar is ready to be used.

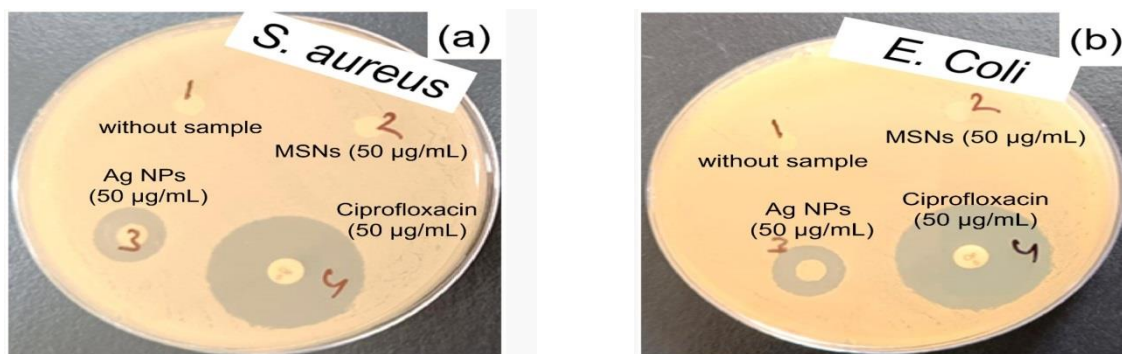
### **4.3.3 Determination of Antibacterial Activity**

The antibacterial activity of silver nanoparticles was evaluated using the standard procedure, the agar disk diffusion technique. After melting and adding 20 ml of the by turning it. This method is referred to as "carpet culture" or "lawn culture." Five minutes of drying the plates is necessary to ensure that the medium fully absorbs the inoculum.

Before handling the antibiotic discs, sterilize the forceps with alcohol. Ideally, discs should be spaced 24 mm apart. To establish proper contact and prevent misplacing, gently touch each disk with the forceps. Plates were covered with paraffin tape and incubated in an incubator at 37°C for around 24 hours. The antibacterial activity was ascertained by measuring the zone of inhibition's millimeter diameter and comparing it to reference values.

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sterile nutritional agar medium to petri plates, it was allowed to cool and harden for half an hour. The inoculum is dipped into a sterile cotton swab, which is then pressed against the tube wall to remove any surplus media. Swipe the whole surface area of the plate.



**Antibacterial activity of silver nanoparticles usin *S.aureus* and *E. coli***

<b><i>S. aureus</i></b>			<b><i>E. Coli</i></b>		
<b>Sample</b>	<b>Concentration</b>	<b>Zone of inhibition</b>	<b>Sample</b>	<b>Concentration</b>	<b>Zone of inhibition</b>
MSNs	---	---	MSNs	---	---
Ag@MSNs	(50 µg/mL)	~12 mm	Ag@MSNs	(50 µg/mL)	~11 mm
Control	(50 µg/mL)	~20 mm	Control	(50 µg/mL)	~20 mm

**Table4.1 shows the zone of inhibition for silver nanoparticles and control ciprofloxacin.**

**CONCLUSION**

Biosynthesis of silver nanoparticles using neem plant extract have been found from this method to produce NPs process high reducing and capping properties, Which are exhibited in the formation of aqueous silver nanoparticle dispersion, through with little variations. They are formed after 5 minutes of mixing silver nitrate. A simple rapid production of silver NPs have been achieved by using neem plant extract. The green production rout is done at room temperature using water and leaves using biological material, therefore neem plant leaf extract was good for the synthesis of silver nanoparticles. X-Ray diffraction spectroscopy was used to

measure the particle size of synthesized silver nanoparticles. XRD confirm the particle size of NPs which is 18.31nm. Uv visible spectroscopy give the absorption of light at 430nm. The most effective antibacterial action was revealed by synthesized AgNPs against human pathogens as *E.coli* and Staphylococcus aureus. Upcoming industrial and medicinal difficulties might be readily met by scaling up the above-mentioned environmentally friendly synthesis process of AgNPs.

## REFERENCES

- Abid, N., Khan, A. M., Shujait, S., Chaudhary, K., Ikram, M., Imran, M., Haider, J., Khan, M., Khan, Q., & Maqbool, M. (2022). Synthesis of nanomaterials using various top-down and bottom-up approaches, influencing factors, advantages, and disadvantages: A review. *Advances in Colloid and Interface Science*, 300, 102597.
- Ahmadi, O., Jafarizadeh-Malmiri, H., & Jodeiri, N. (2018). Eco-friendly microwave-enhanced green synthesis of silver nanoparticles using Aloe vera leaf extract and their physico-chemical and antibacterial studies. *Green Processing and Synthesis*, 7(3), 231-240.
- Alahmad, A., Al-Zereini, W. A., Hijazin, T. J., Al-Madanat, O. Y., Alghoraibi, I., Al-Qaralleh, O., Al-Qaraleh, S., Feldhoff, A., Walter, J.-G., & Scheper, T. (2022). Green synthesis of silver nanoparticles using Hypericum perforatum L. aqueous extract with the evaluation of its antibacterial activity against clinical and food pathogens. *Pharmaceutics*, 14(5), 1104.
- Alsammarraie, F. K., Wang, W., Zhou, P., Mustapha, A., & Lin, M. (2018). Green synthesis of silver nanoparticles using turmeric extracts and investigation of their antibacterial activities. *Colloids and Surfaces B: Biointerfaces*, 171, 398-405.
- Asif, M., Yasmin, R., Asif, R., Ambreen, A., Mustafa, M., & Umbreen, S. (2022). Green Synthesis of Silver Nanoparticles (AgNPs), Structural Characterization, and their Antibacterial Potential. *Dose-Response*, 20(2), 15593258221088709.
- Balamurugan, M., & Saravanan, S. (2017). Green synthesis of silver nanoparticles by using Eucalyptus Globulus leaf extract. *Journal of The Institution of Engineers (India): Series A*, 98, 461-467.
- Banerjee, P., Satapathy, M., Mukhopahayay, A., & Das, P. (2014). Leaf extract mediated green synthesis of silver nanoparticles from widely available Indian plants: synthesis, characterization, antimicrobial property and toxicity analysis. *Bioresources and Bioprocessing*, 1, 1-10.
- CG Kiruba Daniel, S., Nehru, K., & Sivakumar, M. (2012). Rapid biosynthesis of silver nanoparticles using Eichornia crassipes and its antibacterial activity. *Current Nanoscience*, 8(1), 125-129.
- Chakravarty, A., Ahmad, I., Singh, P., Sheikh, M. U. D., Aalam, G., Sagadevan, S., & Ikram, S. (2022). Green synthesis of silver nanoparticles using fruits extracts of Syzygium cumini and their bioactivity. *Chemical Physics Letters*, 795, 139493.
- Chand, K., Abro, M. I., Aftab, U., Shah, A. H., Lakhani, M. N., Cao, D., Mehdi, G., & Mohamed, A. M. A. (2019). Green synthesis characterization and antimicrobial activity

- against *Staphylococcus aureus* of silver nanoparticles using extracts of neem, onion and tomato. *RSC advances*, 9(30), 17002-17015.
- Deivanathan, S. K., & Prakash, J. T. J. (2022). Green synthesis of silver nanoparticles using aqueous leaf extract of *Guettarda Speciosa* and its antimicrobial and anti-oxidative properties. *Chemical Data Collections*, 38, 100831.
  - Golabiazar, R., Othman, K. I., Khalid, K. M., Maruf, D. H., Aulla, S. M., & Yusif, P. A. (2019). Green synthesis, characterization, and investigation antibacterial activity of silver nanoparticles using *Pistacia atlantica* leaf extract. *Bionanoscience*, 9(2), 323-333.
  - Gour, A., & Jain, N. K. (2019). Advances in green synthesis of nanoparticles. *Artificial cells, nanomedicine, and biotechnology*, 47(1), 844-851.
  - Hawar, S. N., Al-Shmgani, H. S., Al-Kubaisi, Z. A., Sulaiman, G. M., Dewir, Y. H., & Rikisahedew, J. J. (2022). Green synthesis of silver nanoparticles from *Alhagi graecorum* leaf extract and evaluation of their cytotoxicity and antifungal activity. *Journal of Nanomaterials*, 2022, 1-8.
  - Hochella Jr, M. F. (2002). Nanoscience and technology: the next revolution in the Earth sciences. *Earth and Planetary Science Letters*, 203(2), 593-605.
  - Jemilugba, O. T., Parani, S., Mavumengwana, V., & Oluwafemi, O. S. (2019). Green synthesis of silver nanoparticles using *Combretum erythrophyllum* leaves and its antibacterial activities. *Colloid and Interface Science Communications*, 31, 100191.
  - Khanal, L. N., Sharma, K. R., Paudyal, H., Parajuli, K., Dahal, B., Ganga, G., Pokharel, Y. R., & Kalauni, S. K. (2022). Green synthesis of silver nanoparticles from root extracts of *Rubus ellipticus* Sm. and comparison of antioxidant and antibacterial activity. *Journal of Nanomaterials*, 2022, 1-11.
  - Khane, Y., Benouis, K., Albukhaty, S., Sulaiman, G. M., Abomughaid, M. M., Al Ali, A., Aouf, D., Fenniche, F., Khane, S., & Chaibi, W. (2022). Green synthesis of silver nanoparticles using aqueous Citrus limon zest extract: Characterization and evaluation of their antioxidant and antimicrobial properties. *Nanomaterials*, 12(12), 2013.
  - Kingsley, J. D., Ranjan, S., Dasgupta, N., & Saha, P. (2013). Nanotechnology for tissue engineering: need, techniques and applications. *journal of pharmacy research*, 7(2), 200-204.
  - Kolahalam, L. A., Viswanath, I. K., Diwakar, B. S., Govindh, B., Reddy, V., & Murthy, Y. (2019). Review on nanomaterials: Synthesis and applications. *Materials Today: Proceedings*, 18, 2182-2190.
  - Kumar, R., Kumar, M., & Luthra, G. (2023). Fundamental approaches and applications of nanotechnology: A mini review. *Materials Today: Proceedings*.
  - K p, F.  .,  oşkunçay, S., & Duman, F. (2020). Biosynthesis of silver nanoparticles using leaf extract of *Aesculus hippocastanum* (horse chestnut): Evaluation of their antibacterial, antioxidant and drug release system activities. *Materials Science and Engineering: C*, 107, 110207.
  - Lalitha, A., Subbaiya, R., & Ponmurugan, P. (2013). Green synthesis of silver nanoparticles from leaf extract *Azadirachta indica* and to study its anti-bacterial and antioxidant property. *Int J Curr Microbiol App Sci*, 2(6), 228-235.
  - Manik, U., Nande, A., Raut, S., & Dhoble, S. (2020). Green synthesis of silver nanoparticles using plant leaf extraction of *Artocarpus heterophyllus* and *Azadirachta indica*. *Results in Materials*, 6, 100086.
  - Mehmood, A., Ghafar, H., Yaqoob, S., Gohar, U. F., & Ahmad, B. (2017). Mesoporous silica nanoparticles: a review. *J. Dev. Drugs*, 6(02).



- Misra, R., Acharya, S., & Sahoo, S. K. (2010). Cancer nanotechnology: application of nanotechnology in cancer therapy. *Drug discovery today*, 15(19-20), 842-850.
- Mody, V. V., Siwale, R., Singh, A., & Mody, H. R. (2010). Introduction to metallic nanoparticles. *Journal of Pharmacy and bioallied sciences*, 2(4), 282.
- Mudhafar, M., Zainol, I., Jaafar, C. N. A., Alsailawi, H., & Majhool, A. A. (2020). Microwave-assisted green synthesis of Ag nanoparticles using leaves of Melia Dubia (Neem) and its antibacterial activities. *Journal of Advanced Research in Fluid Mechanics and Thermal Sciences*, 65(1), 121-129.
- Nazeruddin, G., Prasad, N., Waghmare, S., Garadkar, K., & Mulla, I. (2014). Extracellular biosynthesis of silver nanoparticle using Azadirachta indica leaf extract and its antimicrobial activity. *Journal of Alloys and Compounds*, 583, 272-277.
- Peralta-Videa, J. R., Huang, Y., Parsons, J. G., Zhao, L., Lopez-Moreno, L., Hernandez-Viezas, J. A., & Gardea-Torresdey, J. L. (2016). Plant-based green synthesis of metallic nanoparticles: scientific curiosity or a realistic alternative to chemical synthesis? *Nanotechnology for Environmental Engineering*, 1, 1-29.
- Rajkumar, P., Prakasam, A., Rajeshkumar, S., Gomathi, M., Anbarasan, P., & Chandrasekaran, R. (2020). Green synthesis of silver nanoparticles using Gymnema sylvestre leaf extract and evaluation of its antibacterial activity. *South African Journal of Chemical Engineering*, 32(1), 1-4.
- Rangasamy, M. (2011). Nano technology: a review. *Journal of applied pharmaceutical science*(Issue), 08-16.
- Rao, B., & Tang, R.-C. (2017). Green synthesis of silver nanoparticles with antibacterial activities using aqueous Eriobotrya japonica leaf extract. *Advances in natural sciences: Nanoscience and nanotechnology*, 8(1), 015014.
- Roy, P., Das, B., Mohanty, A., & Mohapatra, S. (2017). Green synthesis of silver nanoparticles using Azadirachta indica leaf extract and its antimicrobial study. *Applied Nanoscience*, 7(8), 843-850.
- Salayová, A., Bedlovičová, Z., Daneu, N., Baláž, M., Lukáčová Bujňáková, Z., Balážová, L., & Tkáčiková, L. (2021). Green synthesis of silver nanoparticles with antibacterial activity using various medicinal plant extracts: Morphology and antibacterial efficacy. *Nanomaterials*, 11(4), 1005.
- Salunke, B. K., Shin, J., Sawant, S. S., Alkotaini, B., Lee, S., & Kim, B. S. (2014). Rapid biological synthesis of silver nanoparticles using Kalopanax pictus plant extract and their antimicrobial activity. *Korean Journal of Chemical Engineering*, 31, 2035-2040.
- Sana, S. S., & Dogiparthi, L. K. (2018). Green synthesis of silver nanoparticles using Givotia moluccana leaf extract and evaluation of their antimicrobial activity. *Materials Letters*, 226, 47-51.
- Senthilkumar, P., Rashmitha, S., Veera, P., Ignatious, C. V., SaiPriya, C., & Samrot, A. V. (2018). Antibacterial activity of neem extract and its green synthesized silver nanoparticles against Pseudomonas aeruginosa. *Journal of Pure and Applied Microbiology*, 12(2), 969-974.
- Sun, Q., Cai, X., Li, J., Zheng, M., Chen, Z., & Yu, C.-P. (2014). Green synthesis of silver nanoparticles using tea leaf extract and evaluation of their stability and antibacterial activity. *Colloids and surfaces A: Physicochemical and Engineering aspects*, 444, 226-231.
- . S. (2016). Controllable synthesis of silver nanoparticles using Neem leaves and their Verma, A., & Mehata, M antimicrobial activity. *Journal of radiation Research and applied sciences*, 9(1), 109-115.

**BIOSYNTHESIS OF MESOPOROUS SILICA SUPPORTED SILVER NANOPARTICLES FOR  
ANTIBACTERIAL EVALUATION**

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- Widatalla, H. A., Yassin, L. F., Alrasheid, A. A., Ahmed, S. A. R., Widdatallah, M. O., Eltilib, S. H., & Mohamed, A. A. (2022). Green synthesis of silver nanoparticles using green tea leaf extract, characterization and evaluation of antimicrobial activity. *Nanoscale Advances*, 4(3), 911-915.
- Yadav, S. G., Patil, S. H., Patel, P., Nair, V., Khan, S., Kakkar, S., & Gupta, A. D. (2018). Green synthesis of silver nanoparticles from plant sources and evaluation of their antimicrobial activity. *Adv. Anal. Tools Mater. Charact*, 5, 133-139.
- Yousaf, H., Mehmood, A., Ahmad, K. S., & Raffi, M. (2020). Green synthesis of silver nanoparticles and their applications as an alternative antibacterial and antioxidant agents. *Materials Science and Engineering: C*, 112, 110901.